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Article

Synthesis and *in Silico* Evaluation of Novel Compounds for PET-Based Investigations of the Norepinephrine Transporter

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Abstract: Since the norepinephrine transporter (NET) is involved in a variety of diseases, the investigation of underlying dysregulation-mechanisms of the norepinephrine (NE) system is of major interest. Based on the previously described highly potent and selective NET ligand 1-(3-(methylamino)-1-phenylpropyl)-3-phenyl-1,3-dihydro-2*H*-benzimidaz- ol-2-one (Me@APPI), this paper aims at the development of several fluorinated methylamine-based analogs of this compound. The newly synthesized compounds were computationally

evaluated for their interactions with the monoamine transporters and represent reference compounds for PET-based investigation of the NET.

Keywords: NET; ADHD; cocaine dependence; BAT; PET; FAPPI

1. Introduction

Abnormal regulation of the norepinephrine transporter (NET) or NET dysfunction, respectively, cause either increased or decreased levels of norepinephrine (NE) in the synaptic cleft. Since NE is a fundamental neurochemical messenger, its accurate regulation is of major importance. Thus, the NET, responsible for NE equilibrium in the synaptic cleft, is representing the reuptake site and considered to be involved in a variety of neurological/psychiatric disorders [1,2], but also plays a pivotal role in cardiovascular [1–3] and metabolic diseases [3–5]. Reduced NET levels go along with neurological disorders like major depression [6,7], Parkinson's disease (PD), Alzheimer's disease (AD) [8–18], and cardiovascular diseases such as hypertension, cardiomyopathy, and heart failure [5,13]. Furthermore, a dysfunction of the NE system was reported in Attention Deficit Hyperactivity Disorder (ADHD) [9,17,19], suicide [1,12,20], substance abuse (cocaine dependence) [16], and schizophrenia [10]. A more recent discovery is the involvement of the NET in diseases like diabetes and obesity, due to its presence in brown adipose tissue (BAT) and the proposed activation thereof via NE [4,5,21].

Based on the fact that the NET is involved in such a variety of diseases, the investigation of the underlying dysregulation-mechanism of the NE system is of major interest. For this purpose, information about the transporter abundance and density in healthy and pathological living human brains is required. The most suitable and accurate technique to gain this information is positron emission tomography (PET). As a non-invasive molecular imaging technique, it represents a suitable approach towards the collection of missing data in the living organism and direct quantification of receptor/transporter densities *in vivo*. To fully gain insight in the molecular changes of the noradrenergic system via PET, however, prior development of suitable NET PET radioligands is required.

So far, radiolabeled NET binding reboxetine analogs [¹¹C]MeNER, [¹¹C]MRB, ((*S*,*S*)-2-(α -(2-[¹¹C]-methoxyphenoxy)benzyl)morpholine) and [¹⁸F]FMeNER-D₂ ((*S*,*S*)-2-(α -(2-[¹⁸F]fluoro[²H₂]methoxyphenoxy)benzyl)morpholine) have been described, which however display certain limitations such as metabolic instability, complex radiosyntheses, or late equilibria [22–26].

Recently, Zhang *et al.* [26] evaluated a series of benzimidazolone-based propanamines with *in vitro* inhibitory activity on the human norepinephrine transporter (hNET). The results of these investigations suggested that compounds containing a phenyl moiety directly attached at the benzimidazolone ring (e.g., **1**, Figure 1) were the most potent, representing a half maximal inhibitory concentration (IC₅₀) below 10 nM (IC₅₀ < 10 nM). Furthermore, hNET selectivity over human serotonin transporter (hSERT) turned out >300-fold superior to those of reboxetine and atomoxetine (16- and 81-fold) [26]. Fluorination at position 2 of the phenyl moiety attached to the benzimidazolone ring (e.g., **1**) and additionally exhibited hNET selectivity over hSERT (80-fold) similar to atomoxetine [26].



Figure 1. Structures of the highly potent and selective NET ligands 1-(3-(<u>methylamino</u>)- 1phenylpropyl)-3-phenyl-1,3-dihydro-2*H*-benzimidazol-2-one (Me@APPI, **1**) and 4-fluoro- 1-(2-fluorophenyl)-3-(3-(methylamino)-1-phenylpropyl)-1,3-dihydro-2*H*-benzimidazol-2one (**2**) as well as radiolabeled analog [¹¹C]Me@APPI (**3**).

Both benzimidazolone derived propanamines with a phenyl moiety (e.g., 1), as well as a fluorinated phenyl moiety (as in 2) indicated excellent hNET selectivity over human dopamine transporter (hDAT) with < 50% inhibition of the cocaine analog [³H]WIN-35,428, binding to hDAT at a concentration of 10 μ M [26]. Given those findings, both described benzimidazolone-based propanamines 1 and 2 represent excellent candidates for selective and potent NET inhibition with high affinity and low unspecific binding. Thus, on the basis of the results of Zhang *et al.* [26] the methylamino moiety of the core compound 1 has been radiolabeled with ¹¹C and tested by our research group [27]. All investigated preclinical parameters, such as affinity, blood brain barrier penetration, lipophilicity, metabolic degradation, and selectivity showed excellent results, thus suggesting suitability of ¹¹C-radiolabeled 1-(3-(methylamino)-1-phenylpropyl)-3-phenyl-1,3-dihydro-2*H*-benzimidazole-2-one ([¹¹C]Me@APPI) (**3**, Figure 1) as a NET radioligand for use in PET.

Due to successful preclinical testing of [¹¹C]Me@APPI (**3**) and given the excellent *in vitro* results of compound **2**, shown by Zhang *et al.* [26] the aim of this paper is the synthesis and docking studies of several fluorinated analogs **4**–**6** of compound **1** (Figure 2) as reference compounds for their later prepared radioactive analogs. All methylamine-derived benzimidazolone derivatives **4**–**6** will be subjected to affinity, selectivity, and lipophilicity studies towards the NET as well as blood brain barrier penetration experiments at the Medical University of Vienna. The most promising NET ligands will then be selected for the development of new, selective PET tracers for the NET and after radiolabeling with both ¹¹C and ¹⁸F, they will be the subject of further experiments.



Figure 2. Chemical structure of envisaged reference compounds 4–6 (FAPPI:1-3).

2. Results and Discussion

The synthesis of reference compounds 4–6 first required the preparation of side chains 11 and 12, as well as core compounds 16–18. After successful preparation, side chain 11 was merged in a condensation reaction with core compound 16, whereas side chain 12 was reacted with core compounds 17 and 18, prior to halogen exchange and substitution with methylamine (Scheme 1).



Reagents and conditions: (i) THF, EtOH, NaBH₄, $-10 \degree C \rightarrow -5 \degree C$, 10 min, yields for 9: 95%, 10: 100%; (ii) aq. HBr, rt, 3 h, yields for 11: 64%, 12: 86%; (iii) 1,1'-carbonyldiimidazole, anhyd. THF, rt, overnight, yields for 16: 74%, 17: 61%, 18: 69%; (iv) 1,1'-carbonyldiimidazole, anhyd. DMF, 90 °C, 2 h, yields for 17: 80%, 18: 80%; (v) K₂CO₃, DMF, rt, 30 min \rightarrow addition of 11 and 12 \rightarrow rt, 30 min, yields for 19: 32%, 20: 63%, 21: 55%; (vi) NaI, acetone, reflux, 24 h, yields for 22: 82%, 23: 76%, 24: 53%; (vii) NH₃ in isopropanol, 80 °C, 3 h, yield for 25: 50%; (viii) methylamine in EtOH, 80 °C, 3 h, yields for 4: 48%, 5: 29%, 6: 30%.

Scheme 1. Synthesis of compounds 4–6.

For the synthesis of side chains **11** and **12**, a protocol of Varney *et al.* [28] was adopted and the keto group of commercially available compounds **7** and **8** was reduced with sodium borohydride in order to

obtain intermediate alcohols 9 and 10. Subsequent bromination of 9 and 10 with aqueous hydrogen bromide led to the formation of products 11 and 12, respectively.

Core compound 16 was prepared by the reaction of commercially available *N*-phenylbenzene-1,2diamine (13) with 1,1'-carbonyldiimidazole. For the preparation of 17 and 18 however, 14 and 15 first had to be made accessible (Scheme 2). Thus, 1-fluoro-2-nitrobenzene (26) reacted with commercially available fluoroanilines 27 and 28, respectively, to obtain disubstituted amines 29 and 30. Therefore, two different methods were applied (Scheme 2): The first method (i) was conducted by heating 26 and 27 with anhydrous potassium fluoride and potassium carbonate in a microwave oven [29]. Since the adoption of an alternative method [30]—conventional heating at 180 °C—gave compound 29 in higher yields, this approach (ii) was chosen for the large scale synthesis of 29 as well as for the preparation of 30.



Reagents and conditions: (i) anhyd. KF, K₂CO₃, microwave oven, 900 W, 10 min, yields for **29**: 58%; (ii) anhyd. KF, K₂CO₃, 180 °C, 2 d, yields for **29**: 68%, **30**: 52%; (iii) Zn, AcOH, 0 °C \rightarrow rt, 2 h, yields for **14**: 93%, **15**: n.d.

Scheme 2. Synthesis of compounds 14–15.

In the next reaction step, the nitro groups of both disubstituted amines **29** and **30** were reduced. For this purpose **29** or **30** were added to a mixture of zinc/acetic acid. The resulting amines **14** or **15** were obtained in excellent yields (Scheme 2) [30].

Freshly prepared intermediates **14** and **15** were then subjected to a cyclization reaction with 1,1'-carbonyldiimidazole in DMF under anhydrous conditions (Scheme 1) by modifying a synthesis protocol according to Zhang *et al.* [26]. DMF was preferred over THF to ensure higher yields and shorter reaction times.

Condensation reactions of benzimidazolones 16, 17, and 18 with side chains 11 and 12, respectively were performed under basic conditions (Scheme 1) by adapting a procedure of Jona *et al.* [31]. After purification, the chloro-substituted derivatives 19–21 were converted into the iodo compounds 22–24 in a Finkelstein reaction. Target compounds 4–6 (FAPPI:1-3) were then obtained by heating derivatives 22–24 in a solution of methylamine in ethanol in a sealed tube for 3 h. In addition to reference compounds 4–6, free amine 25 was synthesized by dissolving 22 in a solution of ammonia in isopropanol and heating the resulting mixture in a sealed tube for 3 h. As compound 25 features a free amine moiety, it can be considered a precursor for radiosynthesis.

Since compounds 4 and 5 comprise a novel fluoro substitution, a computational docking study was performed to assess if these compounds still would fit in the binding site of the NET. Furthermore, we aimed at creating a binding mode hypothesis which allows gaining insights into the molecular basis of binding and selectivity towards the monoamine transporters. As the basic scaffold has been shown to act in

an enantioselective manner, the respective (*R*) enantiomers were used throughout the docking studies [26]. The ligands were docked in the substrate binding site (S1) of the outward-open conformation of the transporter models (see Experimental Section for details), since related inhibitors, such as nortriptyline, sertraline, mazindol, *etc.* were also shown to fit in the S1 of the *Drosophila* DAT (dDAT) and the "SERT"-ized leucine transporter ("LeuBAT") in the same protein conformation [32,33]. Interestingly, the co-crystallized ligand in dDAT, nortriptyline (**31**), has the same ranking of human NET, SERT and DAT activity as reference compound **1**, *i.e.*, 4.4, 18 and 1149 nM K_D *vs.* 9, 2995 and >10,000 nM IC₅₀, respectively [26,34]. Additionally, nortriptyline shares important structural features with the benzimidazolones, *i.e.*, two aromatic moieties and an *N*-methyl-ethylenamine side chain. Therefore their binding mode can be expected to be similar.

Common scaffold clustering [35] revealed two binding hypotheses (see Experimental Section) which indicated that compounds **4–6** fit in the S1 of all three transporters. Hence, additional fluorination does not seem to cause steric clashes. In both hypotheses, the most prominent protein-ligand interaction was the cationic nitrogen atom placed in the A sub pocket [36], located between the central Asp75/98/79 side chain as a salt-bridge and the Phe72/95/76 side chain as a cation-*pi* interaction in NET/SERT/DAT, respectively. This is well in accordance with the X-ray structures of the templates. Additional *pi-pi* stacking interactions with Phe152/176/156 and Phe323/355/341 further promote the binding in both hypotheses obtained:

Hypothesis 1: the benzimidazolone heterocycle (ring 2) is placed in the B sub pocket and ring 3 in the C pocket, whereas ring 1 is solvent exposed (see figure in Experimental Section).

Hypothesis 2: Ligand ring 1 is placed in the B pocket whereas ring 2 is placed at the same height (measured from the membrane-water interface) and overlap with the rings of nortriptyline (**31**, Figure 3). The solvent exposed Tyr151/Tyr175/Phe155 in NET/SERT/DAT, resp. T-stacks with ring 2 whereas ring 3 points extracellularly (Table 1, Figure 4).



Figure 3. Chemical structure of nortriptyline (co-crystallized ligand from the template, PDB code 4M48) [28].

Table 1. Sequence alignment of all distinct monoamine transporter residues located in sub sites (reference [32]) in the vicinity of the docked compounds. Red: hydrophilic side chain, green: lipophilic side chain, bold: bulkier side chain.

B Site				C Site			Outer Site		
hNET	S420	M424	G149	V148	A145	F72	D473	Y151	A477
hSERT	T439	L443	A173	I172	A177	Y95	E493	Y175	T497
hDAT	A423	M427	G153	V152	S149	F76	D476	F155	A480



Figure 4. Overlay of compounds **1**, **2**, and **4–6** (maroon) in binding hypothesis 2 in hNET showing agreement with the co-crystal pose of nortriptyline (**31**) (green). Val148 and Asp473 allow more space than Ile172 and Glu439 in hSERT, resp., whereas Tyr151 might induce a more potent stacking interaction as compared to Phe155 in hDAT. The angle between ligand ring 2 and 3 is almost 90° in all poses. The extracellular space is located above.

As binding hypothesis 2 is in close agreement with the co-crystallized ligands in dDAT and LeuBAT, we focus further analysis on this proposed binding mode. Binding hypothesis 2 indicates why the investigated compounds (4–6) show weaker affinity to SERT and DAT than to NET: lower SERT affinity may be due to Ile172 and Glu439, allowing less space for the ligand to be accommodated as compared to in NET, that comprises a valine and an aspartate at the homologous positions, respectively.

Lower DAT affinity could be ascribed to weaker T stacking interactions of Phe155 as compared to Tyr151 in NET, based on previous findings that a Tyr-Phe pair has a stronger binding energy than a Phe-Phe pair [37].

Since the docking studies indicate that fluorinated methyl amines **4–6** (FAPPI:1-3) bind in an analogous way to the NET as reference compound **1** [26], compounds **4–6** will be employed in future studies and evaluated for affinity and selectivity towards the NET. Additionally, lipophilicity studies and blood brain barrier penetration experiments are planned for compounds **4–6** at the Medical University of Vienna. The most promising derivatives regarding their suitability as NET ligands will then be selected for the further development of new and selective PET tracers for the NET, which will comprise either a [¹¹C]methylamine, [¹⁸F]fluoroalkyl amine or [¹⁸F]fluorobenzene radiolabel, respectively. The results of ongoing studies on affinity, selectivity and lipophilicity of the discussed compounds, will be published in a subsequent paper.

3. Experimental Section

3.1. General

The NMR spectra were recorded from CDCl₃ or DMSO-*d*₆ solutions on a Bruker DPX200 spectrometer (200 MHz for ¹H, 50 MHz for ¹³C) or on a Bruker Avance III 400 spectrometer (400 MHz

for ¹H, 100 MHz for ¹³C, 40 MHz for ¹⁵N, 376 MHz for ¹⁹F) at 25 °C. The center of the solvent (residual) signal was used as an internal standard which was related to TMS with δ 7.26 ppm (¹H in CDCl₃), δ 2.49 ppm (¹H in DMSO-*d*₆), δ 77.0 ppm (¹³C in CDCl₃) and δ 39.5 ppm (¹³C in DMSO-*d*₆). ¹⁵N NMR spectra (gs-HMBC, gs-HSQC) were referenced against neat, external nitromethane, ¹⁹F NMR spectra by absolute referencing via Ξ ratio. Digital resolutions were 0.25 Hz/data point in the ¹H and 0.3 Hz/data point in the ¹³C-NMR spectra. Coupling constants (*J*) are quoted in Hz. The following abbreviations were used to show the multiplicities: s: singlet, d: doublet, t: triplet, q: quadruplet, dd: doublet of doublet, m: multiplet. Mass spectra were obtained on a Shimadzu QP 1000 instrument (EI, 70 eV), high-resolution mass spectrometry (HRMS) was carried out on a Finnigan MAT 8230 (EI, 10 eV) or Finnigan MAT 900 S (ESI, 4 kV, 3 µA, CH₃CN/MeOH) electrospray ionization mass spectrometer with a micro-TOF analyzer. Microwave experiments were carried out in a Synthos 3000 microwave oven (SXQ80 rotor, Anton Paar, Graz, Austria) with an internal temperature probe. Compound purity: all compounds synthesized featured a purity of at least 95%.

3.2. Syntheses

3.2.1. General Procedure for the Synthesis of 9 and 10

Starting materials 7 or 8, respectively (1 mmol) was dissolved in THF (1 mL) and EtOH (1 mL) was added. The mixture was cooled to -10 °C and NaBH₄ (1.05 mmol) was slowly added at this temperature. The solution was stirred at -5 °C for 10 min and thereafter, poured into a mixture of saturated aqueous ammonium chloride (3 mL) in ice (1.5 g). The product was extracted with diethyl ether, dried over Na₂SO₄ and evaporated to dryness. The crude product was employed directly in the subsequent reaction step without further purification.

3-Chloro-1-(4-fluorophenyl)propan-1-ol (**9**). Yield: 4.78 g (95%), pale yellow oil, analytical data are in complete accordance with literature values [38].

3-Chloro-1-phenylpropan-1-ol (**10**). Yield: 4.61 g (99%), light yellow oil, analytical data are in complete accordance with literature values [28].

3.2.2. General Procedure for the Synthesis of 11 and 12

To starting material 9 or 10, respectively (1 mmol) was added 48% aqueous HBr (3 mL) and the mixture was stirred for 3h at room temperature. Thereafter, the solution was poured into a mixture of K_2CO_3 (1 g) in ice (5.5 g) and additional solid K_2CO_3 was added for neutralization (pH 7). The crude reaction product was extracted with diethyl ether, the combined organic layers were dried with MgSO₄ and evaporated to dryness. The crude product was employed directly in the subsequent reaction step without further purification.

1-(1-Bromo-3-chloropropyl)-4-fluorobenzene (11). Yield: 64%, pale yellow oil, analytical data are in complete accordance with literature values [39].

(1-Bromo-3-chloropropyl)benzene **(12)**. Yield: 5.64 g (86%), yellow oil, analytical data are in complete accordance with literature values [28].

3.2.3. General Procedure for the Synthesis of 29 and 30

4-Fluoroaniline or 2-fluoroaniline, respectively (1 mmol), anhydrous KF (1 mmol), and K₂CO₃ (1 mmol) were well powdered with a mortar and a pestle, then 1-fluoro-2-nitrobenzene (1 mmol) was added and the mixture was stirred for 2 days at 180 °C. Thereafter, water (5 mL) and CH₂Cl₂ (5 mL) were added and the organic layer was washed with 10% HCl (5 mL) and brine (5 mL). The combined organic layers were dried over Na₂SO₄ and evaporated to dryness prior to purification by column chromatography.

N-(4-Fluorophenyl)-2-nitroaniline (**29**). Yield: 68%, dark orange crystals, mp. 82–83 °C, purification: silica gel 60, petroleum ether/ethyl acetate 9:1 and RP-18 silica gel, methanol/water 7:3, analytical data are in complete accordance with literature values [29].

2-Fluoro-N-(2-nitrophenyl)aniline (**30**). Yield: 52%, orange crystals, mp. 79–80 °C, purification: silica gel 60, petroleum ether/ethyl acetate 9:1, analytical data are in complete accordance with literature values [40].

3.2.4. Alternative Procedure for the Synthesis of 29

4-Fluoroaniline (7.89 g, 6.73 mL, 70.87 mmol), anhydrous KF (4.13 g, 70.87 mmol), and K₂CO₃ (9.81 g, 70.87 mmol) were well powdered with a mortar and a pestle, then 1-fluoro-2-nitrobenzene (10.00 g, 7.47 mL, 70.87 mmol) was added and the mixture was irradiated in the microwave (900 W, 10 min). Thereafter, water (8 mL) and CH₂Cl₂ (10 mL) were added and the organic layer was washed with 10% HCl (5 mL) and brine (5 mL). The combined organic layers were dried over Na₂SO₄ and evaporated to dryness prior to purification by column chromatography (silica gel 60, petroleum ether/ethyl acetate 9.5:0.5). Yield: 9.51 g (58%), dark orange crystals, mp. 82 °C–83 °C.

3.2.5. General Procedure for the Synthesis of 14 and 15

To a solution of Zn^0 (13.8 mmol) in glacial acetic acid (1 mL) was added starting material **28** or **29** (1 mmol) at 0 °C under argon atmosphere. After the addition, the mixture was allowed to warm to room temperature and was stirred for 2 h. Zn^0 was filtered off and the pH of the solution was adjusted to pH 9 with 2N NaOH. Thereafter, the aqueous layer was extracted three times with CH₂Cl₂, the combined organic layers were dried over MgSO₄ and evaporated to dryness.

N-(4-Fluorophenyl)benzene-1,2-diamine (14). Yield: 93%, dark orange-reddish oil, purification: silica gel 60, petrol ether/ethyl acetate 9:1, analytical data are in complete accordance with literature values [30].

N-(2-Fluorophenyl)benzene-1,2-diamine (15). The crude reaction product was subjected to the next reaction step without further purification.

3.2.6. General Procedure for the Synthesis of 16-18

To a solution of starting materials **13**, **14**, or **15** (1 mmol) in THF was added 1,1'-carbonyldiimidazole (1.4 mmol) under argon atmosphere and the mixture was stirred at room temperature overnight. Thereafter, the crude reaction product was purified by column chromatography.

1-Phenyl-1,3-dihydro-2H-benzimidazol-2-one (**16**). Yield: 0.85 g (74%), pink crystals, mp. 201 °C–202 °C, THF: 10 mL, purification: silica gel 60, petroleum ether/ethyl acetate 1:1, analytical data are in complete accordance with literature values [41].

1-(4-Fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one (**17**). Yield: 61%, brown resin, THF: 25 mL, purification: silica gel 60, petroleum ether/ethyl acetate 9:1, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 6.74–6.81 (m, 2H), 6.89–7.11 (m, 2H), 7.14–7.23 (m, 2H), 7.48–7.59 (m, 1H), 7.73–7.78 (m, 1H), 9.03 (br s, 1H), ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 108.5, 110.0, 116.3, 116.8, 121.4, 121.7, 122.3, 128.0, 128.2, 130.4, 135.1, 155.1, 159.3, 164.2, MS: *m/z* (%) 228 (M⁺, 100), 199 (31), 172 (8), 114 (9), 95 (10), 75 (17), 51 (10), HRMS: *m/z* calculated for C₁₃H₁₀FN₂O [M + H]⁺: 229.0772. Found: 229.0769.

1-(2-Fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one (**18**). Yield: 69%, brown resin, THF: 20 mL, purification: silica gel 60, petroleum ether/ethyl acetate 9:1, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 6.82–6.85 (m, 1H), 7.00–7.19 (m, 3H), 7.26–7.88 (m, 2H), 7.43–7.60 (m, 2H), 10.54 (br s, 1H), ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 108.9, 110.1, 117.0, 117.4, 121.5, 121.9, 122.4, 124.9, 125.0, 128.2, 129.6, 130.4, 154.8, 155.4, 160.5, MS: *m/z* (%) 228 (M⁺, 33), 199 (9), 181 (15), 149 (17), 111 (22), 97 (20), 71 (41), 69 (100), 55 (53), 43 (56), HRMS: *m/z* calculated for C₁₃H₉FN₂NaO [M + Na]⁺: 251.0597. Found: 251.0592.

3.2.7. Alternative Procedure for the Synthesis of 17 and 18

A solution of 1,1'-carbonyldiimidazole (1.4 mmol) in DMF (4 mL) was slowly added to a mixture of **14** or **15** (1 mmol) in DMF (4 mL) under argon atmosphere. The resulting solution was stirred at 90 °C for 2 h. After completion of the reaction, the solvent was evaporated *in vacuo*, the slurry was taken up in water, filtered and dried.

1-(4-Fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one (17). Yield: 80%, brown resin.

1-(2-Fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one (18). Yield: 80%, brown resin.

3.2.8. General Procedure for the Synthesis of 19-21

Starting materials 16–18 (1 mmol) and K_2CO_3 (2 mmol) were suspended in DMF (1.8 mL) and stirred at 25 °C for 30 min. 11 and 12, respectively (1.5 mmol) were added after 30 min and the solution was stirred at room temperature overnight. To the mixture was added ethyl acetate (5 mL) and water (5 mL). The aqueous layer was extracted several times with ethyl acetate (10 mL) and the combined organic layers were washed with brine, dried over MgSO₄ and evaporated to dryness.

1-(3-Chloro-1-(4-fluorophenyl)propyl)-3-phenyl-1,3-dihydro-2H-benzimidazol-2-one (**19**). Yield: 32%, white oil, purification: silica gel 60, petroleum ether/ethyl acetate 8:2, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.70–2.87 (m, 1H), 3.12–3.30 (m, 1H), 3.60–3.66 (m, 2H), 5.73 (m, 1H), 7.00–7.11 (m, 6H), 7.38–7.57 (m, 7H), ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 34.3, 42.0, 53.8, 108.7, 109.0, 115.5, 115.9, 121.7, 122.0, 126.0, 127.8, 129.2, 129.4, 129.5, MS: *m/z* (%) 380 (M⁺, 21), 210 (100) (M⁺-C₉H₉ClF), 181 (8), 167 (12), 135 (9), 115 (5), 109 (58), 77 (12), HRMS: *m/z* calculated for C₂₂H₁₈ClFN₂ONa [M + Na]⁺: 403.0989. Found: 403.0989.

I-(*3*-*Chloro-1-phenylpropyl)-3-(4-fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one* (**20**). Yield: 63%, dark orange resin, purification: silica gel 60, petroleum ether/ethyl acetate 9:1 and RP-18 silica gel, methanol, ¹H-NMR (400 MHz, CDCl₃): δ (ppm) 2.80–2.88 (m, 1H, 2'-C<u>H</u>₂), 3.17–3.26 (m, 1H, 2'-C<u>H</u>₂), 3.62–3.70 (m, 2H, 3'-C<u>H</u>₂), 5.79 (dd, J = 10.0 Hz and 5.6 Hz, 1H, 1'-C<u>H</u>), 7.04–7.09 (m, 4H, benzim 4-C<u>H</u>, benzim 5-C<u>H</u>, benzim 6-C<u>H</u>, benzim 7-C<u>H</u>), 7.21–7.26 (m, 2H, f-phen 3-C<u>H</u>, f-phen 5-C<u>H</u>), 7.31–7.33 (m, 1H, phen 4-C<u>H</u>), 7.36–7.40 (m, 2H, phen 3-C<u>H</u>, phen 5-C<u>H</u>), 7.52–7.56 (m, 4H, f-phen 2-C<u>H</u>, f-phen 6-C<u>H</u>, phen 2-C<u>H</u>, phen 6-C<u>H</u>), 1³C-NMR (100 MHz, CDCl₃): δ (ppm) 34.1 (2'-CH₂), 42.0 (3'-C₂H₂), 54.4 (1'-C₂H), 108.6 (benzim 4-C₂H), 109.0 (benzim 7-C₂H), 116.4 (d, J = 22.9 Hz, f-phen 3-C₂H), 121.6 (benzim 5-C₂H), 122.1 (benzim 6-C₂H), 127.4 (phen 2-C₂H), 127.4 (phen 6-C₂H), 127.9 (d, J = 8.6 Hz, f-phen 2-C₂H), 127.9 (d, J = 8.6 Hz, f-phen 2-C₂H), 127.9 (d, J = 8.6 Hz, f-phen 3-C₂H), 128.1 (phen 4-C<u>H</u>), 128.7 (benzim 7a-C), 128.8 (phen 3-C₂H), 128.8 (phen 5-C₂H), 129.4 (benzim 3a-C), 130.3 (d, J = 3.1 Hz, f-phen 1-C), 138.4 (phen 1-C), 153.2 (benzim 2-C₂O), 161.6 (d, J = 247.7 Hz, f-phen 4-C₂F), ¹⁹F-NMR (471 MHz, CDCl₃): δ (ppm) -113.31 (m, 5-C<u>F</u>), MS: *m/z* (%) 380 (M⁺, 2), 228 (100) (M⁺-C₉H₁₀Cl), 199 (11), 185 (16), 153 (6), 117 (14), 91 (73), 75 (8), HRMS: *m/z* calculated for C₂₂H₁₉ClFN₂O [M + H]⁺: 381.1170. Found: 381.1176.

1-(3-Chloro-1-phenylpropyl)-3-(2-fluorophenyl-1,3-dihydro-2H-benzimidazol-2-one (**21**). Yield: 55%, orange resin, purification: silica gel 60, petroleum ether/ethyl acetate 9:1, ¹H-NMR (400 MHz, CDCl₃): δ (ppm) 2.78–2.86 (m, 1H, 2'-C<u>H</u>₂), 3.23 (br s, 1H, 2'-C<u>H</u>₂), 3.64–3.68 (m, 2H, 3'-C<u>H</u>₂), 5.79 (br s, 1H, 1'-C<u>H</u>), 6.85–6.87 (m, 1H, benzim 4-C<u>H</u>) 7.05–7.06 (m, 3H, benzim 5-C<u>H</u>, benzim 6-C<u>H</u>, benzim 7-C<u>H</u>), 7.28–7.40 (m, 5H, f-phen 3-C<u>H</u>, f-phen 6-C<u>H</u>, phen 3-C<u>H</u>, phen 4-C<u>H</u>), phen 5-C<u>H</u>), 7.43–7.48 (m, 1H, f-phen 4-C<u>H</u>), 7.52–7.56 (m, 3H, f-phen 5-C<u>H</u>, phen 2-C<u>H</u>, phen 6-C<u>H</u>), ¹³C-NMR (100 MHz, CDCl₃): δ (ppm) 34.2 (2'-CH₂), 41.9 (3'-CH₂), 54.5 (1'-C₂H), 108.8 (d, *J* = 1.7 Hz, benzim 4-C₂H), 109.0 (benzim 7-C<u>H</u>), 117.1 (d, *J* = 19.5 Hz, f-phen 3-C<u>C</u>H), 121.6 (benzim 5-C<u>H</u>), 121.9 (f-phen 1-C), 122.0 (benzim 6-C<u>C</u>H), 124.8 (d, *J* = 3.9 Hz, f-phen 6-C<u>C</u>H), 127.3 (phen 6-C<u>C</u>H), 128.1 (phen 4-C<u>C</u>H), 128.8 (phen 3-C<u>C</u>H), 128.8 (phen 3-C<u>C</u>H), 129.4 (benzim 3a-C), 129.5 (f-phen 5-C<u>C</u>H), 130.2 (d, *J* = 7.8 Hz, f-phen 4-C<u>C</u>H), 138.4 (phen 1-C), 153.0 (benzim 2-C<u>O</u>), 157.9 (d, *J* = 253.2 Hz, f-phen 2-C<u>C</u>F), due to limited resolution of the measuring apparatus, quaternary carbon benzim 7a-C could not be detected, ¹⁹F-NMR (471 MHz, CDCl₃): δ (ppm) -118.39 (m, f-phen 2-C<u>F</u>), MS: *m/z* (%) 380 (M⁺, 12), 228 (100) (M⁺-C₉H₁₀Cl), 199 (5), 153 (3), 117 (7), 91 (49), 75 (5), HRMS: *m/z* calculated for C₂₂H₁₉ClFN₂O [M + H]⁺: 381.1170. Found: 381.1164.

3.2.9. General Procedure for the Synthesis of 22-24

A solution of starting materials **19**, **20**, or **21** (1 mmol) and NaI (1.03 g, 6.89 mmol) in acetone (7 mL) was refluxed for 24 h. The precipitate formed was filtered and the solvent was removed *in vacuo*.

1-(1-(4-Fluorophenyl)-3-iodopropyl)-3-phenyl-1,3-dihydro-2H-benzimidazol-2-one (**22**). Yield: 82%, yellow resin, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.77–2.92 (m, 1H), 3.14–3.31 (m, 3H), 5.63–5.70 (m, 1H), 7.01–7.10 (m, 6H), 7.36–7.56 (m, 7H), ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 2.3, 35.3, 57.0, 108.8, 108.9, 115.5, 115.9, 121.6, 122.0, 126.0, 127.7, 128.5, 129.1, 129.3, 129.4, 134.0, 134.1, 134.3, 153.0, 159.9, 164.8, MS: *m/z* (%) 472 (M⁺, 32), 317 (8), 210 (100) (M⁺-C₉H₉F), 181 (11), 167 (23), 140 (3), 135 (43), 115 (8), 109 (34), 77 (15), 51 (7), HRMS: *m/z* calculated for C₂₂H₁₈FIN₂ONa [M + Na]⁺: 495.0346. Found: 495.0353.

1-(4-Fluorophenyl)-3-(3-iodo-1-phenylpropyl)-1,3-dihydro-2H-benzimidazol-2-one (23). Yield: 76%, yellow crystals, mp. 39 °C–41 °C, purification: silica gel 60, petrol ether/ethyl acetate 9:1, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.81–2.99 (m, 1H, 2'-C<u>H</u>₂), 3.11–3.35 (m, 3H, 2'-C<u>H</u>₂, 3'-C<u>H</u>₂), 5.70 (dd, J = 6 Hz and 2 Hz, 1H, 1'-C<u>H</u>), 7.04–7.09 (m, 4H, benzim 4-C<u>H</u>, benzim 5-C<u>H</u>, benzim 6-C<u>H</u>, benzim 7-C<u>H</u>), 7.17–7.42 (m, 5H, f-phen 3-C<u>H</u>, f-phen 5-C<u>H</u>, phen 4-C<u>H</u>, phen 2-C<u>H</u>, phen 5-C<u>H</u>), 7.51–7.57 (m, 4H, f-phen 2-C<u>H</u>, f-phen 6-C<u>H</u>, phen 2-C<u>H</u>, phen 6-C<u>H</u>), 108.6 (benzim 4-C<u>H</u>), 109.2 (benzim 7-C<u>H</u>), 116.4 (d, J = 23 Hz, f-phen 5-C<u>H</u>), 121.6 (benzim 5-C<u>H</u>), 122.1 (benzim 6-C<u>H</u>), 127.3 (phen 2-C<u>H</u>), 127.8 (phen 4-C<u>C</u>H), 128.1 (d, J = 5 Hz, f-phen 2-C<u>H</u>), 128.5 (benzim 7a-C), 128.8 (phen 3-C<u>C</u>H), 128.8 (phen 5-C<u>C</u>H), 129.4 (benzim 3a-C), 130.3 (d, J = 3Hz, f-phen 1-C), 138.1 (phen 1-C), 153.2 (benzim 2-CO), 161.6 (d, J = 247 Hz, f-phen 4-C<u>C</u>F), MS: m/z (%) 472 (M⁺, 13), 317 (5), 228 (100) (M⁺-C₉H₁₀I), 199 (8), 185 (15), 117 (47), 103 (2), 91 (40), 75 (8), 55 (5), HRMS: m/z calculated for C₂₂H₁₉FIN₂O [M + H]⁺: 473.0526. Found: 473.0506.

1-(2-Fluorophenyl)-3-(3-iodo-1-phenylpropyl)-1,3-dihydro-2H-benzimidazol-2-one (**24**). Yield: 53%, yellow crystals, mp. 38 °C–39 °C, purification: silica gel 60, petrol ether/ethyl acetate 9:1, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.80–2.98 (m, 1H, 2'-C<u>H</u>₂), 3.13–3.36 (m, 3H, 2'-C<u>H</u>₂, 3'-C<u>H</u>₂), 5.67–5.74 (m, 1H, 1'-C<u>H</u>), 6.85–6.88 (m, 1H, benzim 4-C<u>H</u>) 6.99–7.07 (m, 3H, benzim 5-C<u>H</u>, benzim 6-C<u>H</u>, benzim 7-C<u>H</u>), 7.26–7.45 (m, 5H, f-phen 3-C<u>H</u>, f-phen 6-C<u>H</u>, phen 3-C<u>H</u>, phen 4-C<u>H</u>, phen 5-C<u>H</u>), 7.47–7.58 (m, 4 H, f-phen 4-C<u>H</u>, f-phen 5-C<u>H</u>, phen 2-C<u>H</u>, phen 6-C<u>H</u>), ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 2.3 (2'-C<u>H</u>₂), 35.4 (3'-C<u>H</u>₂), 57.6 (1'-C<u>H</u>), 108.9 (d, *J* = 1.5 Hz, benzim 4-C<u>C</u>H), 109.1 (benzim 7-C<u>H</u>), 117.1 (d, *J* = 19 Hz, f-phen 3-C<u>H</u>), 121.6 (benzim 5-C<u>H</u>), 121.9 (f-phen 1-C), 122.0 (benzim 6-C<u>C</u>H), 124.8 (d, *J* = 3.5 Hz, f-phen 6-C<u>C</u>H), 127.3 (phen 2-C<u>C</u>H), 127.3 (phen 4-C<u>C</u>H), 128.1 (phen 4-C<u>C</u>H), 128.8 (phen 3-C<u>C</u>H), 129.5 (f-phen 5-C<u>C</u>H), 130.2 (d, *J* = 8 Hz, f-phen 4-C<u>C</u>H), 138.2 (phen 1-C), 153.0 (benzim 2-<u>C</u>O), 157.8 (d, *J* = 251.5 Hz, f-phen 2-<u>C</u>F), MS: *m/z* (%) 472 (M⁺, 9), 317 (5), 241 (4), 228 (100) (M⁺-C₉H₁₀I), 199 (5), 185 (10), 117 (37), 91 (29), 75 (7), HRMS: *m/z* calculated for C₂₂H₁₉FIN₂O [M + H]⁺: 473.0526. Found: 473.0532.

3.2.10. General procedure for the synthesis of *1-(3-amino-1-(4-fluorophenyl)propyl)-3-phenyl-1,3-dihydro-2H-benzimidazol-2-one* (**25**)

1-(1-(4-fluorophenyl)-3-iodopropyl)-3-phenyl-1,3-dihydro-2*H*-benzimidazol-2-one (0.26 g, 0.55 mmol) and a solution of NH₃ in isopropanol (2 M, 22 mL) were heated in a sealed tube for 3 h at 80 °C. After evaporation of the solvent, the crude product was purified by column chromatography (silica gel 60, CH₂Cl₂/MeOH 9:1). Yield: 0.10 g (50%), light brown crystals, mp. 87–88 °C. ¹H-NMR (400 MHz, CDCl₃): δ (ppm) 2.74–2.84 (m, 3H, 2'-CH₂, 3'-CH₂), 2.98–3.04 (m, 1H, 3'-CH₂), 5.74–5.78 (m, 1H, 1'-CH), 6.79-6.81 (m, 1H, benzim 7-CH), 6.96-7.01 (m, 5H, benzim 4-CH, benzim 5-CH, benzim 6-CH, f-phen 3-CH, f-phen 5-CH), 7.30-7.33 (m, 1H, phen 4-CH), 7.40-7.48 (m, 4H, f-phen 2-CH, f-phen 6-CH, phen 3-CH, phen 5-CH), 7.52-7.54 (m, 2H, phen 2-CH, phen 6-CH), due to limited resolution of the instrumentation, the NH₂ protons could not be detected, ¹³C-NMR (100 MHz, CDCl₃): δ (ppm) 30.3 (2'-<u>C</u>H₂), 37.8 (3'-<u>C</u>H₂), 52.7 (1'-<u>C</u>H), 109.2 (benzim 4-<u>C</u>H), 110.0 (benzim 7-CH), 115.7 (d, J = 21.5 Hz, f-phen 3-CH), 115.7 (d, J = 8.2 Hz, f-phen 5-CH), 122.0 (benzim 5-CH), 122.3 (benzim6-CH), 126.4 (phen 2-CH), 126.4 (phen 6-CH), 127.5 (benzim 7a-C), 128.1 (phen 4-CH), 129.2 (d, J = 8.2 Hz, f-phen 2-CH), 129.2 (d, J = 8.2 Hz, f-phen 6-CH), 129.5 (benzim 3a-C), 129.7 (phen 3-CH), 129.7 (phen 5-CH), 133.3 (d, J = 3.2 Hz, f-phen 1-C), 134.0 (phen 1-CH), 153.8 (benzim 2-CO), 162.3 (d, J = 247.4 Hz, f-phen 4-<u>C</u>F), ¹⁹F-NMR (471 MHz, CDCl₃): δ (ppm) -113.68 (m, f-phen C<u>F</u>), MS: m/z(%) 361 (M⁺, 17), 331 (10), 210 (100) (M⁺-C₉H₁₁FN), 181 (15), 167 (16), 149 (29), 128 (17), 77 (19), 57 (20), 43 (12), HRMS: m/z calculated for C₂₂H₂₁FN₃O [M + H]⁺: 362.1669. Found: 362.1674.

3.2.11. General Procedure for the Synthesis of 4-6

Starting materials **18**, **19** or **20** (1 mmol) and a solution of methylamine in EtOH (12.5 mL, 8 M) were heated in a sealed tube for 3 h at 80 °C. After evaporation of the solvent, the crude reaction product was purified by column chromatography.

1-(1-(4-Fluorophenyl)-3-(methylamino)propyl)-3-phenyl-1,3-dihydro-2H-benzimidazol-2-one (4). Yield: 48%, light orange resin, purification: silica gel 60, dichloromethane/methanol 9:1 and dichloromethane/ethyl acetate/methanol 7:2:1, ¹H-NMR (400 MHz, CDCl₃): δ (ppm) 2.42 (s, 3H, NHCH₃), 2.57–2.74 (m, 4H, 2'-CH₂, 3'-CH₂), 3.15 (br s, 1H, NHCH₃), 5.76–5.79 (m, 1H, 1'-CH), 6.88–6.90 (m, 1H, benzim 7-CH), 6.97–7.05 (m, 4H, benzim 5-CH, benzim 6-CH, f-phen 3-CH, f-phen 5-CH), 7.07-7.10 (m, 1H, benzim 4-CH), 7.39-7.43 (m, 1H, phen 4-CH), 7.46-7.57 (m, 6H, f-phen 2-CH, f-phen 6-CH, phen 2-CH, phen 3-CH, phen 5-CH, phen 6-CH), ¹³C-NMR (100 MHz, CDCl₃): δ (ppm) 30.6 (2'-CH₂), 35.9 (NHCH₃), 48.3 (3'-CH₂), 53.1 (1'-CH), 108.9 (benzim 4-CH), 109.5 (benzim 7-CH), 115.5 (d, J = 21.5 Hz, f-phen 3-CH), 115.5 (d, J = 21.5 Hz, f-phen 5-CH) 121.4 (benzim 5-CH), 121.8 (benzim 6-CH), 126.0 (phen 2-CH), 126.0 (phen 6-CH), 127.7 (phen 4-CH), 128.0 (benzim 7a-C), 129.0 (d, J = 8.1 Hz, f-phen 2-CH), 129.0 (d, J = 8.1 Hz, f-phen 6-CH), 129.4 (benzim 3a-C), 129.5 (phen 3-CH), 129.5 (phen 5-C<u>H</u>), 134.4 (phen 1-<u>C</u>H), 134.6 (d, J = 3.4 Hz, f-phen 1-<u>C</u>H), 153.5 (benzim 2-<u>C</u>O), 162.1 (d, J = 246.7 Hz, f-phen 4-<u>C</u>F), ¹⁹F-NMR (471 MHz, CDCl₃): δ (ppm) -114.36 (m, f-phen 4-C<u>F</u>), MS: *m*/*z* (%) 375 (M⁺, 16), 210 (57) (M⁺-C₁₀H₁₃FN), 181 (10), 167 (12), 150 (10), 109 (22), 97 (16), 71 (27), 57 (78), 44 (100), HRMS: *m/z* calculated for C₂₃H₂₃FN₃O [M + H]⁺: 376.1825. Found: 376.1821.

1-(4-Fluorophenyl)-3-(3-(methylamino)-1-phenylpropyl)-1,3-dihydro-2H-benzimidazol-2-one (5). Yield: 29%, light yellow crystals, mp. 100 °C–102 °C, purification: silica gel 60, dichloromethane/methanol 9:1 and RP-18 silica gel methanol/water 9:1 and 7:3, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.53 (s, 3H, NHCH₃), 3.13 (br s, 4H, 2'-CH₂, 3'-CH₂), 5.77–5.81 (m, 1H, 1'-CH), 6.96-7.03 (m, 4H, benzim 4-CH, benzim 5-CH, benzim 6-CH, benzim 7-CH), 7.16-7.34 (5H, f-phen 3-CH, f-phen 5-CH, phen 4-CH, phen 2-CH, phen 5-CH), 7.49-7.56 (m, 4H, f-phen 2-CH, f-phen 6-CH, phen 2-CH, phen 6-CH), due to limited resolution of the instrumentation, the NH proton could not be detected, ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 27.6 (2'-CH₂), 33.1 (NHCH₃), 47.0 (3'-CH₂), 54.2 (1'-CH), 108.8 (benzim 4-CH), 109.9 (benzim 7-CH), 116.6 (d, J = 23 Hz, f-phen 3-CH), 116.6 (d, J = 23 Hz, f-phen 5-CH), 122.1 (benzim 5-CH), 122.6 (benzim 6-CH), 127.2 (phen 2-CH), 127.2 (phen 6-<u>C</u>H), 127.7 (phen 4-<u>C</u>H), 128.3 (d, J = 6 Hz, f-phen 2-<u>C</u>H), 128.3 (d, J = 6 Hz, f-phen 6-<u>C</u>H), 128.4 (benzim 7a-C), 128.9 (phen 3-CH), 128.9 (phen 5-CH), 129.2 (benzim 3a-C), 129.7 (d, J = 3 Hz, f-phen 1-C), 136.9 (phen 1-C), 153.4 (benzim 2-CO), 161.7 (d, J = 248 Hz, f-phen 4-CF), MS: m/z (%) 375 (M⁺, 11), 330 (7), 228 (50) (M⁺-C₁₀H₁₄N), 199 (7), 185 (9), 147 (17), 128 (26), 117 (8), 91 (13), 58 (34), 44 (100), HRMS: *m/z* calculated for C₂₃H₂₃FN₃O [M + H]⁺: 376.1825. Found: 376.1828.

1-(2-Fluorophenyl)-3-(3-(methylamino)-1-phenylpropyl)-1,3-dihydro-2H-benzimidazol-2-one **(6)**. Yield: 30%, yellow crystals, mp. 92 °C-93 °C, purification: silica gel 60, dichloromethane/methanol 9:1 and RP-18 silica gel methanol/water 9:1 and 7:3, ¹H-NMR (200 MHz, CDCl₃): δ (ppm) 2.57 (s, 3H, NHCH₃), 3.01–3.15 (m, 4H, 2'-CH₂, 3'-CH₂), 5.75–5.86 (m, 1H, 1'-CH), 6.83–6.87 (m, 1H, benzim 4-CH) 6.98-7.05 (m, 3H, benzim 5-CH, benzim 6-CH, benzim 7-CH), 7.25-7.42 (m, 5H, f-phen 3-CH, f-phen 6-CH, phen 3-CH, phen 4-CH, phen 5-CH), 7.48–7.61 (m, 4 H, f-phen 4-CH, f-phen 5-CH, phen 2-CH, phen 6-CH), due to limited resolution of the instrumentation, the NHCH₃ proton could not be detected, ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) 27.7 (2'-<u>C</u>H₂), 33.2 (NH<u>C</u>H3), 46.9 (3'-<u>C</u>H₂), 53.5 (1'-<u>C</u>H), 109.0 (benzim 4-CH), 110.1 (benzim 7-CH), 117.1 (d, J = 19.5 Hz, f-phen 3-CH), 122.2 (benzim 5-CH), 122.7 (benzim 6-CH), 125.1 (d, J = 3.0 Hz, f-phen 6-CH), 127.3 (phen 2-CH), 127.1 (phen 6-CH), 128.3 (phen 4-CH), 128.9 (phen 3-CH), 128.9 (phen 5-CH), 129.5 (f-phen 5-CH), 130.7 (d, J = 4.5 Hz, f-phen 4-CH), 136.5 (phen 1-C), 153.6 (benzim 2-CO), 157.6 (d, J = 250.5 Hz, f-phen 2-CF), due to limited resolution of the measuring apparatus, quaternary carbon f-phen 1-C could not be detected, MS: m/z (%) 375 (M⁺, 17), 318 (10), 228 (82) (M⁺-C₁₀H₁₄N), 199 (9), 185 (9), 147 (16), 128 (35), 117 (9), 91 (20), 58 (43), 44 (100), HRMS: m/z calculated for C₂₃H₂₃FN₃O [M + H]⁺: 376.1825. Found: 376.1822.

3.3. Computational Methods

The ligand structures were built in the protonated form using Molecular Operating Environment (MOE) 2013 [42]. Homology models of human NET, SERT and DAT were obtained from the *Drosophila* dopamine transporter template (dDAT_{cryst}, PDB id 4M48 [32]), by selecting the model with the most favorable Discrete Optimized Protein Energy (DOPE) of 250 generated by Modeller 9.11 [43]. The co-crystallized inhibitor nortriptyline was retained during model generation and the compounds were docked in the same site using Genetic Optimization for Ligand Docking (GOLD) 5.2 [44]. One hundred poses per ligand (*i.e.*, five hundred poses per protein target) were generated based on the GoldScore scoring function, while keeping the ligand flexible and the protein rigid.

The common chemical scaffold, *i.e.*, the reference compound, was extracted from the resulting poses, analogous to the methods of our previous study [35]. Cluster analysis was performed based on Euclidian distance and complete linkage of the root-mean square deviation of the ligand's heavy atoms matrix using XLStat [45]. The dendrogram was cut at eight clusters and the ones containing all five ligands were selected (Figure 5).



Figure 5. *Left column*: Overlay of compounds **1**, **2**, and **4–6** (maroon) in binding hypothesis 1 and comparison with the co-crystal pose of nortriptyline (**31**) (green). In NET, V148 allows more space than I172 in SERT. The angle between ligand ring 2 and 3 is *ca*. 60° in all poses. *Right column*: Overlay of compounds **1**, **2**, and **4–6** (maroon) in binding hypothesis 2. In NET, V148 still allows more space than in hSERT, whereas E439 might disrupt ligand ring 3. DAT lacks a more potent stacking interaction due to F155 as compared to NET[Y151] and SERT[Y175]. Binding mode 2 poses are more in agreement with the co-crystal pose. The angle between ligand ring 2 and 3 is almost 90° in all poses. The extracellular space is located above in all figures.

4. Conclusions

In conclusion, ten new compounds have been synthesized within the scope of this work, which aimed at the development of new, selective, high affinity references for the imaging of the NET system via PET. Four of these new compounds (4–6 and 25) will be employed in future studies. Whilst methylamines 4–6 (FAPPI:1-3) represent reference compounds for their later prepared radioactive analogs, additionally prepared free amine 25 (APPI:1) will serve as precursor for radiolabeling. Since docking studies indicate that fluorinated methyl amines 4–6 (FAPPI:1-3) bind in an analogous way to the NET as reference compound 1, these compounds 4–6 are promising candidates for biological evaluation.

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Author Contributions

Catharina Neudorfer: Responsible for the performance of the syntheses and writing; Amir Seddik: Structure activity relationships calculations; Karem Shanab: Contributions to syntheses and experimental procedures; Andreas Jurik: Structure activity relationships calculations; Christina Rami-Mark: Participated in design and performance of the experiments; Wolfgang Holzer: Performance of the NMR analyses; Gerhard Ecker: Conceived and supervised the SAR experiments; Markus Mitterhauser: Designed parts of the research and proofread the manuscript; Wolfgang Wadsak: Designed parts of the research and proofread the manuscript; Helmut Spreitzer: Conceived and supervised the syntheses.

Conflicts of Interest

The authors declare no conflict of interest.

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Sample Availability: Samples of the compounds 4–6, 9–25, and 29 and 30 are available from the authors.

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